The functional morphology of 'the tennis wrist'

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Abstract

At present there is no consensus in the literature concerning the aetiology of lateral epicondylitis and a causal treatment. Microscopic lesions in the common origin of wrist and finger extensors, which are caused by excessive strain, are thought to be the underlying cause of lateral epicondylitis. At least 40 different courses of treatment have been described. However, it is still unknown which treatment is considered the best. In 1988, Riezebos and Lagerberg published about the tennis wrist from a functional morphology perspective (Riezebos & Lagerberg 1988).

Several findings, which cannot be explained by excessive strain, are understandable within the concept of lateral epicondylitis caused by an internal derangement of the wrist. The aim of this review is to falsify the model of lateral epicondylitis, referred to as 'the tennis wrist', which has prevailed for more than 20 years.

(key-words: tennis elbow, tendinopathy, etiology, musculoskeletal manipulations, wrist)

Introduction

Lateral epicondylitis (LE) has been clinically diagnosed (Goguin & Rush 2003; Boisaubert et al 2004) on the basis of pain around the epicondyl. The pain either occurs or increases during resistance to the extension of the wrist or 3rd metacarpal (Struijs et al 2003; Hong et al 2004) and the palpation of the lateral epicondyl (Radpasand 2009; Assendelft et al 2009). Gripping is painful, certainly when force is required. Mundane activities, such as shaking hands and holding a cup, become difficult (Trudel et al 2004; Vicenzino et al 2007).

LE has been diagnosed as the most common complaint in elbow problems (Hong et al 2004). In 80% of the cases, the symptoms are chronic (Radpasand 2009). Statistical data reveal that LE can be associated with long-term absenteeism from work (Hong et al 2004; Bisset et al 2005). A recovery period of 6 months to 2 years has been reported (Smidt et al 2002; Coombes et al 2009).

The aetiology of LE is unknown (Slater et al 2003; Manchanda & Grover 2007; Alizadehkhaiyat et al 2009). The underlying cause of lateral epicondylitis is thought to be attributed to microscopic lesions in the common origin of the wrist and finger extensors caused by excessive strain (Benjamin et al 2006; Manchanda & Grover 2007; Cutlip et al 2009; Magnusson et al 2010), especially in the extensor carpi radialis brevis muscle (ECRB) (Alfredson et al 2000; Boisaubert et al 2004; Alizadehkhaiyat et al 2009). At least 40 different types of treatment have been described (Bisset et al 2005; Stasinopoulos & Johnson 2007; Radpasand 2009). However, it remains unclear which treatment is the best.

The first article about LE was published in 1873 (Goguin & Rush 2003; Hong et al 2004). Since then various pathophysiological hypotheses have been described (Snijders et al 1987; Stoeckart et al 1991; Hägg & Milerad 1997).

In 1988, Riezebos and Lagerberg published about the tennis wrist from a functional morphological perspective (Riezebos & Lagerberg 1988). LE could be regarded as the functional morphological changes that occur in the ECRB resulting from a scapholunate internal derangement (Frankel et al 1971; Tasker & Waugh 1982; Dimitroulis 2005) during wrist extension.

The starting point for this article is the 'expiry date' of the controversial principle of 'the tennis wrist', which has prevailed for the past 20 years. For manual therapy, investigating the possibilities of a functional morphological approach is important, as the *functional* morphology enables intervention in the causal function problems underlying the symptoms. The manual therapist then treats the primary disrupted function and not the secondary 'abnormal' form or 'diseased' structure.

Sub-questions

- 1. Is an inflammatory infiltrate found in tendon tissue? Does the tendon tissue undergo morphological changes in LE?
- 2. Do these changes occur on the basis of an altered mechanical strain in the tendon?
- 3. Can an altered ratio rolling/gliding in the joint lead to an altered mechanical strain in the tendon?
- 4. Can an altered positioning of the movement axes in joints lead to an altered ratio rolling/gliding?

Method

The database search was carried out in August, 2010. Using specific search strings, PubMed searched for articles related to the functional morphological changes that occur in tendon tissue as a result of an internal derangement in the spanned joint.

The MeSH search terms used are: 'tendons', 'tendon injuries', 'tennis elbow', 'adaptation, physiological' and the free search term, 'wrist'.

Inclusion criteria

Articles meeting the following criteria have been included:

- 1. Publications from the past 15 years (arbitrary).
- 2. Publications preferably about humans.
- 3. Consistent with the aforementioned sub-questions.
- 4. Publications in English, French, German and Dutch.

Studies at all levels of evidence are eligible for this literature study. The publications included have been rated according to a citation analysis, the journal impact factor and the H index.

Figure 1.

Evaluation 1

The first evaluation takes place on the basis of the research objective.

Evaluation 2

After having studied the full-text version, those articles that appeared to be inconsistent with the subquestions, were excluded.

Results

The search resulted in 157 articles, which have been subjected to further assessment. Furthermore, references found in the literature have been screened and evaluated for their relevance to the research objective. Of the 180 original articles, 63 were excluded after the first evaluation, which means that 117 articles were included for the second evaluation. After the second evaluation, 102 articles remained.

The morphology of epicondylitis

Histopathological research has demonstrated that few macrophages, lymphocytes or neutrophils are found in chronically-overstrained tendon tissue (Benjamin et al 2006; Magnusson et al 2010). The histopathological findings for tendinopathy are characterised by the rupturing and thinning of collagen fibres, increased vascularisation, granulation tissue and an increase in proteoglycans (PG) in the extracellular matrix (Benjamin et al 2006; Riley 2008).

The pathological changes are described as 'a-functional'. It would therefore seem better to talk about 'tendinosis' (Goguin & Rush 2003; Slater et al 2003; Doschak & Zernicke 2005).

Angiofibroblastic tendinosis refers to degenerative changes that occur when a tendon is repeatedly subjected to microtrauma (Kjaer 2004; Boisaubert et al 2004; Coombes et al 2009).

In tendinopathy, molecular changes exhibit an increased expression of type III collagen, decorin, aggrecan and biglycan. These changes indicate recovery processes and adaptive responses to changes in mechanical strain (Hoffmann & Gross 2007; Riley 2008; Lui et al 2010). Decorin and biglycan are 'small leucine-rich repeated proteoglycans' (SLRP) that contribute to collagen fibril structure, whereas aggrecan is a large highly hydrophilic PG that plays a role in adaptation to compressive forces (Riley 2008; Lui et al 2010).

The mechanics of the tendon and the enthesis

The tendon consists of hierarchical layers in which Type I crimped collagen fibres are accurately arranged in the pulling orientation (Yoon & Halper 2005; Riley 2008).

The tendon responds to mechanical strain by altering its structure, composition and mechanical properties. This process is termed 'mechanotransduction'. Intermolecular, covalent cross-linking ensures mechanical stabilization of collagen fibrils and the ultimate tension-absorbing properties of the tendon (Chiquet et al 2003; Martin et al 2003; Screen et al 2006).

Figure 2.

A strong correlation exists between fibre orientation of collagen connective tissue and the functional requirements imposed upon the tissue. If force transfer of tissue is anisotropic, then the functional properties of the resulting structure are as well (Ottani et al 2001; Kjaer et al 2006). In an aponeurotic tendon the force transfer occurs in a non-uniform manner (Maganaris 2002; Thomopoulos et al 2006), which can lead to intratendinous shear strain (Magnusson et al 2008; Benjamin et al 2008; Burkholder 2008).

However, the enthesis is adapted to the effects of non-uniform force transfer as a consequence of an insertional angle change. A correlation exists between the quantity of fibrous cartilage in entheses and the extent to which an insertional angle change occurs during movements (Benjamin et al 2006). In

'fibrocartilaginous entheses' (FCE), chondrogenesis has taken place (Milz et al 2005; Riley 2008; Thomopoulos et al 2010).

Figure 3.

Fibrous cartilage facilitates the dissemination of tension at the area of transition between hard and soft tissue so that the transition is resilient to compression and shear forces. This characteristic is reflected in the molecular composition, namely the presence of specific hyaline cartilage molecules, in particular sulphated GAGs, aggrecan and Type II collagen (Milz et al 2005; Thomopoulos et al 2010). Tendons, which have been subjected to compression forces, develop cartilaginous characteristics (Kraushaar & Nirschl 1999; Ottani et al 2001).

Pathological changes are found at the very locations where 'strain', i.e. elongation, occurs the least. A clinically recognisable enthesopathy mainly occurs in the deeply situated parts of an enthesis where such regions are compressed by the superficial parts, and where the presence of fibrous cartilage is usually more apparent (Benjamin et al 2006; Coombes et al 2009).

The mechanics of epicondylitis

The origin of the ECRB is located on the movement axis of the elbow (London 1981; Lieber et al 1997b). With its relatively short fibres and large moment arm, the ECRB acts as a mechanical torque for wrist extension. This design results in an ECRB that is isometrically strong (Lieber et al 1997a; Lieber & Fridén 1998).

Electromyographic investigations reveal that the ECRB exhibits more electrical activity than the extensor carpi radialis longus muscle (ECRL) (Tournay & Paillard 1953; Riek et al 2000) during extension and gripping. In patients with LE, the ECRB exhibits an increased electrical activity compared to a healthy control group (Bauer & Murray 1999; Shetty & Randolph 2000).

An increase or decrease in the activity of the wrist extensors and flexors following electrical stimulation of the scapholunate interosseous ligament (SLIL) is observed, dependent on the wrist position. Increased activity is encountered in the antagonists of wrist positions and may possibly have a protective function for the joint (Hagert 2010).

The arthrokinematics of epicondylitis

In vivo it has been found that the scaphoid rotates more upon flexion and extension than the lunate, which implies intercarpal rotations (Kobayashi et al 1997; Sun et al 2000; Wolfe et al 2000; Moojen 2003). Intra-articular kinematics are described using arthrokinematic terminology such as rolling, gliding and spinning (Biscević et al 2005; Baeyens et al 2006). Rolling and gliding take place simultaneously (Schomacher 2009).

Joints have incongruent profiles and subsequently no fixed rotational axis (Lagerberg 2007). The instant centre of rotation (ICR) therefore shifts in the joint during movement (Gertzbein et al 1986; Sears et al 2006; Schomacher 2009). For the joint to move, the ICR should always be normal to the contact point. With this, each tendency of the rotational axis to shift is accompanied by a shift of the contact point. Any displacement of the ICR away from the perpendicular of the tangent in the **point of joint contact** leads to compression or separation of the profiles (Montgomery et al 1998).

The direction in which the moment arms of ligaments exert force on the profiles accurately determines the position of the centrode (Herzog & Read 1993; Leardini 2001; Biscević et al 2005). Abnormal positions of a centrode lead to changes in the rolling/gliding ratio (Frankel et al 1971) and vice-versa. Movement-impaired joints exhibit a reduced glide and an increased roll between the surfaces (Schomacher 2009). A joint limitation develops when the direction of the speed of movement on the surfaces has the tendency to compress both profiles. The movement blocks and further flexion or extension is no longer possible (Frankel et al 1971; Montgomery et al 1998).

The SLIL is the principle stabilizing structure between the scaphoid and the lunate (Moojen et al 2002; Kai et al 2006; Milz et al 2006; Short et al 2007). Ligament damage of the SLIL is the most common cause of carpal instability and is a condition that eventually results in severe impairment of radiocarpal function (Blevens et al 1989; Milz et al 2006). After section of the SLIL, normal articular contacts are altered, the contribution of the scaphoid increases and that of the lunate decreases during flexion and extension

(Blevens et al 1989; Tang et al 2002; Kai et al 2006; Henry 2008) and the scapholunate movement becomes variable (Werner et al 2005).

A kinematic change of the scaphoid after rupture of the SLIL alters the position of the surrounding tendons and their centre of rotation, which leads to a change in tendon excursion (Tang et al 1997). Immobilisation also results in instability caused by the ligaments being more compliant, resulting in ligamentous laxity (Wren et al 2000; Doschak & Zernicke 2005; Woo et al 2006; Magnusson et al 2008).

Discussion

The concept of a scapholunate internal derangement being a cause of LE makes several findings comprehensible, which cannot be explained by excessive strain (Boisaubert et al 2004; Magnusson et al 2008; Cutlip et al 2009).

The selective involvement of the ECRB in LE (Slater et al 2003; Manchanda & Grover 2007; Puchner 2008) and the increased electrical activity this muscle exhibits (Bauer & Murray 1999; Shetty & Randolph 2000) are mentioned frequently. In 'the tennis wrist' model, a scapholunate internal derangement is assumed to be responsible for a meticulously described sequence of events.

In this scapholunate internal derangement, the rolling component of the scaphoid becomes too large during extension (Crisco JJ et al 1999; Wolfe et al 2000; Kai et al 2006; Henry 2008), which could lead to a subluxation with increased capsular tension in the complex. As compensation, the ECRB is activated to tilt the 3rd radius (3rd metacarpal, capitate, lunate) as a 'block' with the scaphoid to reduce the associated ligamentous tension (Riezebos & Lagerberg 1988).

In arthrokinematics, it is assumed that the convexity has a 'natural' tendency to roll in the absence of ligamentous guidance (Schomacher 2009). 'The tennis wrist' model also assumes that an internal derangement results in an increased arthrokinematic roll as the glide appears later in the movement phase.

However, in view of the low friction coefficient of joint cartilage, it is questionable whether the rolling component will occur at an unguided moment. When ligamentous guidance is lacking, the position of the ICR is undefined. The ICR defies all arthrokinematic laws, albeit it briefly. It no longer needs to be on the perpendicular of the tangent in the contact point. In short, the ICR is subject to the capriciousness of the resultant outcome of all moments acting on the joint.

In the model, the increased capsular tension in the scapholunate complex results in pain and a response of the ECRB. When electrically stimulating the SLIL, Hagert found either an increase or a decrease in the activity of the wrist extensors and flexors, depending on the wrist position (Hagert 2010). In this study, no distinction was made between the ECRL and the ECRB. However, it does illustrate the tendency of the body to defend itself against the potential threat of end positions.

London positions the rotational axis of the elbow through the centre of the arch formed by the trochlear sulcus and the humeral capitellum (London 1981). Thus the origin of the ECRB falls on the movement axis of the elbow. The mechanical effect of a muscle is determined by the distance from the line of action of the muscle to the ICR. The moment arm that the ECRB has over the elbow is negligible in that sense. This implies that the ECRB fulfills its function during gripping irrespective of the elbow position. With its relatively short fibres and large moment arm, the ECRB provides a large mechanical torque for wrist extension, resulting in an ECRB that is isometrically strong (Lieber et al 1997a; Lieber & Fridén 1998). From these publications, it can be deduced that the ECRB is suitable for isometric loads, as is generally the case during gripping.

If excessive strain can be held responsible for LE, the questions still remain as to why the symptoms do not quickly disappear during rest (Hong et al 2004; Manchanda & Grover 2007; Alizadehkhaiyat et al 2009; Assendelft et al 2009), and why symptoms are found in patients who seldom perform activities requiring considerable exertion (Goguin & Rush 2003; Milz et al 2004).

According to the 'tennis wrist' model, immobility causes internal derangement. This explains why patients, who do not perform physically-demanding activities, can be affected (Riezebos & Lagerberg

1988; Goguin & Rush 2003; Milz et al 2004). Immobility or a 'lack of use' is a very dangerous stimulus to the musculoskeletal system. Immobility results in a 'weaker' connective tissue (Woo et al 1987; Akeson et al 1987; Yasuda & Hayashi 1999; Woo et al 2006; Magnusson et al 2008; Puchner 2008) and subsequently to less ligamentous guidance of the joint. The tendency of a joint with an internal derangement to subluxate results in excessive capsular tension and movement impairment.

In this functionally disrupted situation, the ECRB is required to function in changed positions (Lieber et al 1997a). Only in specific positions can the operating force be evenly distributed over the collagen fibres. In positions other than 'known ones', 'shearing moments' occur in the flat origin (Riezebos & Lagerberg 1988). There is external evidence for the existence of tension concentrations referred to in the 'tennis wrist' model.

The direction (Wang 2006; Magnusson et al 2008) of the forces in the aponeurotic origin of the ECRB no longer concur with the direction in which collagen fibres are oriented (Ottani et al 2002). The force transfer takes place out-of-plane (Thomopoulos et al 2006), resulting in changes in intratendinous shear strain (Benjamin et al 2006; Magnusson et al 2008; Burkholder 2008).

There are indications in the literature for the remodelling proposed in the model.

The histopathological findings for tendinopathy are characterised by 'angiofibroblastic hyperplasia' (Boisaubert et al 2004; Coombes et al 2009). There is evidence for the presence of fibrous cartilage as a normal characteristic of entheses (Benjamin & Ralphs 1998; Milz et al 2004).

Aggrecan plays a role in the adaptation to compressive load (Doschak & Zernicke 2005; Riley 2008; Lui et al 2010). The molecular composition of various fibrous connective tissues is directly related to the respective mechanical tissue functions (Milz et al 2005).

It would seem that tissue in the tendon, ideally suited to resisting tension from one single direction, gradually transforms into tissue that needs to be resilient against tensional forces from various different directions. Consequently, it must also be resilient to compressive forces that arise from deformation of the tissue. Type I collagen connective tissue, with fibres accurately arranged at micrometer level, is replaced by Type III collagen and cartilage.

It is unlikely that 'the body' replaces mechanically-specific tissue without there being a clear mechanical need for this, since remodelling (Ottani et al 2001; Wang 2006), requires a high metabolic activity. The formation of cartilage in the tendon should therefore be seen as a normal, non-diseased development in response to compressive load.

Other than Riezebos and Lagerberg's publication (Riezebos & Lagerberg 1988), nothing else has ever been written about 'the tennis wrist'. Therefore, the main focus has been on factors relevant to this topic. As the model assumes a functional adaptation of the ECRB, the MeSH term, 'adaptation', was used. This might have resulted in selection bias.

As there was no scoring method for descriptive studies, the publications included were evaluated by means of a citation analysis, the journal impact factor and the H index.

The higher the score for one or more ratings, the more authoritative the reference.

Interpretation and proposal

If ligamentous guidance is reduced, an internal derangement occurs as a result of which altered (scapholunate) kinematics develop (Montgomery et al 1998; Brownhill et al 2006).

The movement in the joint becomes erratic and unpredictable (Weiler & Bogoch 1995; Scarvell et al 2005; Werner et al 2005; Foumani et al 2009).

A condition for an equilibrium in the joint is that all moments cancel each other out and yield a reaction force without a resulting joint moment. This 'zero joint reaction moment' can only proceed through the ICR (Pierce & Li 2005; Li et al 2006; Lagerberg 2007).

The friction coefficient in a synovial joint is about 0.01 (Carter et al 2007; Ateshian 2009). In comparison, the friction coefficient of ice on ice is 0.1. Force transfer between profiles can therefore only occur perpendicularly. If the reaction force deviates by a single degree, the profiles would immediately slip (Lagerberg 2007). Ligaments guarantee the alignment of the reaction force without exerting a moment on the joint (Lagerberg 2007; Kleipool & Blankevoort 2010). In cases of ligament injury or immobility, the

contact points proceed on the basis of the prevailing load conditions, resulting in the joint becoming a highly unpredictable mechanical linkage (Lagerberg 2007).

The tendon is confronted with tension concentrations that in terms of direction and intensity are variable and unpredictable (Pierce & Li 2005; Thomopoulos et al 2010).

The fibre orientation in the tendon no longer satisfies the requirements imposed on it. It therefore undergoes a remodelling (Ottani et al 2001; Riley 2008).

Cartilage is resilient against variable tension concentrations due to variable shear and compression forces (Benjamin et al 2006; Lui et al 2010).

Chondrogenesis and osteogenesis are the result of functional morphological adaptations and are not pathologic (Benjamin & Ralphs 1998; Milz et al 2005; Li et al 2010).

In the preliminary stages of calcific tendinitis, the tendon transforms into fibrocartilagenous tissue due to the metaplasia of tenocytes into chondrocytes (Hughes & Bolton-Maggs 2002; Lam et al 2006). The tendon becomes infiltrated with an extracellular matrix of cartilage when it is subjected to compressive loading (Ottani et al 2001).

Calcified tissue contributes to the functional recovery of the enthesis (Newsham-West et al 2007). Intratendinous calcifications and enthesophytes are found in LE (Levin et al 2005). Therefore, the changes that take place in LE cannot be termed 'degenerative' or 'a-functional'. The tendon is actively developing under the prevailing variable mechanical conditions to create tissue that is resilient to the 'freaky' tension in the tendon. Even when forces in tendon tissue are opposing and, as such, extremely conflicting demands are made on it, the tendon transforms into a structure where the 'average' function is integrated.

The changes that occur in the tendon can be described as 'tendopoiesis'. The suffix poiesis means 'construction' and 'development'.

Conclusions

- No inflammatory infiltrate is found in tendon tissue in LE. In tendinopathy, tendon tissue undergoes morphological changes to complete the remodelling.
- 2. These changes can take place in response to changes in the mechanical load.
- 3. These changes in the mechanical strain of the tendon can result from an altered ratio rolling/ gliding in the spanned joint.
- 4. An altered position of the ICR results in an altered rolling/gliding ratio.

The origin of the ECRB undergoes functional morphological changes in LE in order to complete a remodelling. The remodelling can possibly be attributed to a scapholunate internal derangement, resulting in highly variable and unpredictable tensions in the origin of the ECRB.

The study has not been able to falsify the model. So therefore, even though it was first proposed more than 20 years ago, the model has not lost its relevance.

A retrospective cohort study into epicondylitis symptoms in patients with scapholunate instability or in patients, who were injured after a fall, would provide more clarity on the aetiology of LE.

According to the model, scapholunate mobility needs to be restored. Arthrokinematic mobilisation techniques required for this do justice to the results of scientific research (Coyle & Robertson 1998), which has revealed that the Type I collagen connective tissue of impaired joints exhibits an increased compliance. We therefore recommend that a randomised controlled trial be performed with a study population of sufficient size (Struijs et al 2003; Boisaubert et al 2004) with a long-term follow-up (Bisset et al 2005; Puchner 2008).

Tendopoeisis is the term that covers the process described in this article.

Figure 1.

Flow chart of the literature search.

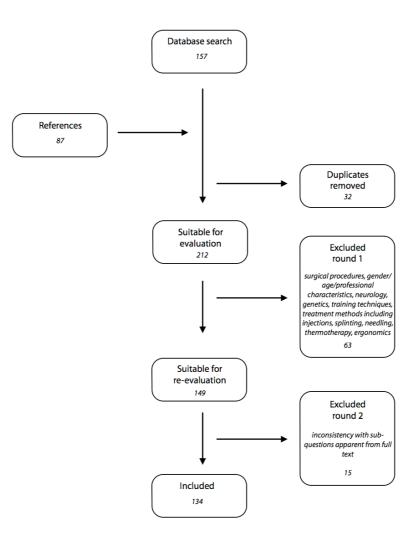
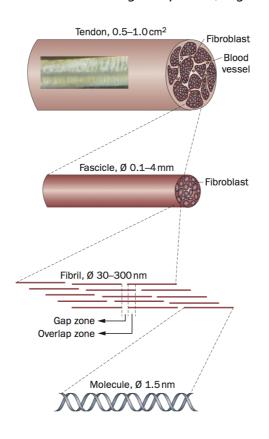


Figure 2.

The various hierarchical levels of tendon tissue. Reproduced by kind permission of Nature Publishing Group from (Magnusson et al 2010).



The tendon is built up from collagen fascicles. The inset shows two bordering collagen fascicles. The crimp pattern of the fascicle is visible.

The fascicle is made up of collagen fibrils, fibroblasts, proteoglycans, glycoproteins and glycosaminoglycans.

The collagen fibril has a 'quarter-stagger-arrangement' of collagen molecules. Crosslinking of the collagen fibrils safeguards the integrity of the fibril.

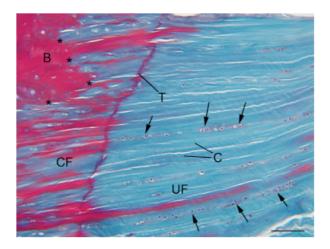
The collagen molecule is built up from three protein chains (α -chains) that form a triple helix.

Figure 3.

Upon approaching the bone (B) the tendon becomes fibrocartilaginous. A section of non-calcified fibrous cartilage (NF) is separated from an area of calcified fibrous cartilage (CF) by a boundary (T). T is the 'hard-soft' boundary of the enthesis. The cells (\rightarrow) in the fibrous cartilage are arranged longitudinally to the collagen fibres (C).

scale line=100 μm.

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References

1. Abbott JH, Patla CE, Jensen RH. The initial effects of an elbow mobilization with movement technique on grip strength in subjects with lateral epicondylalgia. Manual therapy 2001; 6(3): 163-169

2. Akeson WH, Amiel D, Abel MF, Garfin SR, Woo S. Effects of immobilization on joints. Clinical Orthopaedics and Related Research 1987; 21928

3. Alfredson H, Ljung BO, Thorsen K, Lorentzon R. In vivo investigation of ECRB tendons with microdialysis technique--no signs of inflammation but high amounts of glutamate in tennis elbow. Acta orthopaedica Scandinavica 2000; 71(5): 475-479

4. Alizadehkhaiyat O, Fisher AC, Kemp GJ, Vishwanathan K, Frostick SP. Assessment of functional recovery in tennis elbow. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology 2009; 19(4): 631-638

5. Andersson T, Eliasson P, Aspenberg P. Tissue memory in healing tendons: short loading episodes stimulate healing. Journal of applied physiology (Bethesda, Md. : 1985) 2009; 107(2): 417-421

6. Arampatzis A, Karamanidis K, Morey-Klapsing G, De Monte G, Stafilidis S. Mechanical properties of the triceps surae tendon and aponeurosis in relation to intensity of sport activity. Journal of biomechanics 2007; 40(9): 1946-1952

7. Assendelft WJJ, Smidt N, Verdaasdonk AL, Dingjan R, Kolnaar BGM. NHG-Standaard Epicondylitis M60. Huisarts Wet. 2009; 3 140-146

8. Ateshian GA, The role of interstitial fluid pressurization in articular cartilage lubrication. Journal of biomechanics 2009; 42(9): 1163-1176

9. Ateshian GA, Hung CT. Patellofemoral joint biomechanics and tissue engineering. Clinical orthopaedics and related research 2005; 43681

10. Baeyens JP, Van Glabbeek F, Goossens M, Gielen J, Van Roy P, Clarys JP. In vivo 3D arthrokinematics of the proximal and distal radioulnar joints during active pronation and supination. Clinical biomechanics (Bristol, Avon) 2006; 21 Suppl 1 S9-12

11. Bauer JA, Murray RD. Electromyographic patterns of individuals suffering from lateral tennis elbow. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology 1999; 9(4): 245-252

12. Benjamin M, Kaiser E, Milz S. Structure-function relationships in tendons: a review. Journal of anatomy 2008; 212(3): 211-228

13. Benjamin M, McGonagle D. Entheses: tendon and ligament attachment sites. Scandinavian journal of medicine & science in sports 2009; 19(4): 520-527

14. Benjamin M, Ralphs JR. Fibrocartilage in tendons and ligaments--an adaptation to compressive load. Journal of anatomy 1998; 193 (Pt 4) 481-494

15. Benjamin M, Toumi H, Ralphs JR, Bydder G, Best TM, Milz S. Where tendons and ligaments meet bone: attachment sites ('entheses') in relation to exercise and/or mechanical load. Journal of anatomy 2006; 208(4): 471-490

16. Biscević M, Tomić D, Starc V, Smrke D. Gender differences in knee kinematics and its possible consequences. Croatian medical journal 2005; 46(2): 253-260

17. Bisset L, Paungmali A, Vicenzino B, Beller E. A systematic review and meta-analysis of clinical trials on physical interventions for lateral epicondylalgia. British journal of sports medicine 2005; 39(7): 411-22; discussion 411-22

18. Blevens AD, Light TR, Jablonsky WS, et al. Radiocarpal articular contact characteristics with scaphoid instability. The Journal of hand surgery 1989; 14(5): 781-790

19. Boer de MD, Maganaris CN, Seynnes OR, Rennie MJ, Narici MV. Time course of muscular, neural and tendinous adaptations to 23 day unilateral lower-limb suspension in young men. The Journal of physiology 2007; 583(Pt 3): 1079-1091

20. Boisaubert B, Brousse C, Zaoui A, Montigny JP. Les traitements non chirurgicaux de la tendinopathie des épicondyliens. Annales de Readaptation et de Medecine Physique 2004; 47(6): 346-355

21. Brownhill JR, Furukawa K, Faber KJ, Johnson JA, King GJ. Surgeon accuracy in the selection of the flexion-extension axis of the elbow: an in vitro study. Journal of shoulder and elbow surgery / American Shoulder and Elbow Surgeons ... [et al.] 2006; 15(4): 451-456

22. Buchanan CI, Marsh RL. Effects of exercise on the biomechanical, biochemical and structural properties of tendons. Comparative biochemistry and physiology. Part A, Molecular & integrative physiology 2002; 133(4): 1101-1107

23. Burkholder TJ, Mechanotransduction in skeletal muscle. Frontiers in bioscience: a journal and virtual library 2008; 12174

24. Carter MJ, Basalo IM, Ateshian GA. The temporal response of the friction coefficient of articular cartilage depends on the contact area. Journal of biomechanics 2007; 40(14): 3257-3260

25. Chiquet M, Renedo AS, Huber F, Flück M. How do fibroblasts translate mechanical signals into changes in extracellular matrix production? Matrix biology : journal of the International Society for Matrix Biology 2003; 22(1): 73-80

26. Coles JM, Chang DP, Zauscher S. Molecular mechanisms of aqueous boundary lubrication by mucinous glycoproteins. Current Opinion in Colloid & Interface Science 2010;

27. Coombes BK, Bisset L, Vicenzino B. A new integrative model of lateral epicondylalgia. British journal of sports medicine 2009; 43(4):252

28. Couppé C, Kongsgaard M, Aagaard P, et al. Habitual loading results in tendon hypertrophy and increased stiffness of the human patellar tendon. Journal of applied physiology (Bethesda, Md. : 1985) 2008; 105(3): 805-810

29. Coyle JA, Robertson VJ. Comparison of two passive mobilizing techniques following Colles' fracture: a multi-element design. Manual therapy 1998; 3(1): 34-41

30. Crisco JJ, McGovern RD, Wolfe SW. In Vivo Scaphoid and Lunate Kinematics in Wrist Flexion and in Extension. 45th Annual Meeting, Orthopaedic Research Society, Anaheim, California 1999;

31. Cutlip RG, Baker BA, Hollander M, Ensey J. Injury and adaptive mechanisms in skeletal muscle. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology 2009; 19(3): 358-372

32. Dimitroulis G, The prevalence of osteoarthrosis in cases of advanced internal derangement of the temporomandibular joint: a clinical, surgical and histological study. International journal of oral and maxillofacial surgery 2005; 34(4): 345-349

33. Doschak MR, Zernicke RF. Structure, function and adaptation of bone-tendon and bone-ligament complexes. Journal of musculoskeletal & neuronal interactions 2005; 5(1): 35-40

34. Foumani M, Strackee SD, Jonges R, et al. In-vivo three-dimensional carpal bone kinematics during flexion-extension and radio-ulnar deviation of the wrist: Dynamic motion versus step-wise static wrist positions. Journal of biomechanics 2009;

35. Frankel VH, Burstein AH, Brooks DB. Biomechanics of internal derangement of the knee. Pathomechanics as determined by analysis of the instant centers of motion. The Journal of bone and joint surgery. American volume 1971; 53(5): 945-962

36. Fuss FK, The ulnar collateral ligament of the human elbow joint. Anatomy, function and biomechanics. Journal of anatomy 1991; 175203

37. Gertzbein SD, Seligman J, Holtby R, et al. Centrode characteristics of the lumbar spine as a function of segmental instability. Clinical orthopaedics and related research 1986; (208): 48-51

38. Goguin JP, Rush FR. Lateral epicondylitis. What is it really?Current Orthopaedics 2003; 17(5): 386-389 39. Hagert E, Proprioception of the wrist joint: a review of current concepts and possible implications on the rehabilitation of the wrist. Journal of hand therapy : official journal of the American Society of Hand Therapists 2010; 23(1): 2-16; quiz 17

40. Hägg GM, Milerad E. Forearm extensor and flexor muscle exertion during simulated gripping work -- an electromyographic study. Clinical biomechanics (Bristol, Avon) 1997; 12(1): 39-43

41. Henry M, Arthroscopic management of dorsal wrist impingement. The Journal of hand surgery 2008; 33(7): 1201-1204

42. Herzog W, Read LJ. Lines of action and moment arms of the major force-carrying structures crossing the human knee joint. Journal of anatomy 1993; 182 (Pt 2) 213-230

43. Hills BA, Butler BD. Surfactants identified in synovial fluid and their ability to act as boundary lubricants. Annals of the rheumatic diseases 1984; 43(4): 641-648

44. Hoffmann A, Gross G. Tendon and ligament engineering in the adult organism: mesenchymal stem cells and gene-therapeutic approaches. International orthopaedics 2007; 31(6): 791-797

45. Hollman JH, Deusinger RH, Van Dillen LR, Matava MJ. Knee joint movements in subjects without knee pathology and subjects with injured anterior cruciate ligaments. Physical therapy 2002; 82(10): 960-972 46. Hong QN, Durand MJ, Loisel P. Treatment of lateral epicondylitis: where is the evidence? Joint, bone, spine : revue du rhumatisme 2004; 71(5): 369-373

47. Hughes PJ, Bolton-Maggs B. Calcifying tendinitis. Current Orthopaedics 2002; 16(5): 389-394 48. Kai S, Yasumoto S, Takahashi S. Accessory Movement of the Lunate during Active Flexion and Extension Motion of the Wrist. Journal of Physical Therapy Science 2006; 18(2): 161-164

49. Ker RF, The implications of the adaptable fatigue quality of tendons for their construction, repair and function. Comparative biochemistry and physiology. Part A, Molecular & integrative physiology 2002; 133(4): 987-1000

50. Kjaer M, Role of extracellular matrix in adaptation of tendon and skeletal muscle to mechanical loading. Physiological reviews 2004; 84(2): 649-698

51. Kjaer M, Magnusson P, Krogsgaard M, et al. Extracellular matrix adaptation of tendon and skeletal muscle to exercise. Journal of anatomy 2006; 208(4): 445-450

52. Kleipool RP, Blankevoort L. The relation between geometry and function of the ankle joint complex: a biomechanical review. Knee surgery, sports traumatology, arthroscopy : official journal of the ESSKA 2010; 18(5): 618-627

53. Kobayashi M, Berger RA, Nagy L, et al. Normal kinematics of carpal bones: a three-dimensional analysis of carpal bone motion relative to the radius. Journal of biomechanics 1997; 30(8): 787-793 54. Kraushaar BS, Nirschl RP. Current Concepts Review-Tendinosis of the Elbow (Tennis Elbow). Clinical Features and Findings of Histological, Immunohistochemical, and Electron Microscopy Studies. The Journal of Bone and Joint Surgery 1999; 81(2):259

55. Krishnan R, Kopacz M, Ateshian GA. Experimental verification of the role of interstitial fluid pressurization in cartilage lubrication. Journal of orthopaedic research : official publication of the Orthopaedic Research Society 2004; 22(3): 565-570

56. Lagerberg A, De interpretatie van het actief- en passief bewegingsonderzoek. Versus, Tijdschrift voor Fysiotherapie 2007; 25(6):

57. Lam F, Bhatia D, Van Rooyen K, de Beer JF. Modern management of calcifying tendinitis of the shoulder. Current Orthopaedics 2006; 20(6): 446-452

58. Leardini A, Geometry and mechanics of the human ankle complex and ankle prosthesis design. Clinical biomechanics (Bristol, Avon) 2001; 16(8): 706-709

59. Leardini A, O'Connor JJ, Catani F, Giannini S. Kinematics of the human ankle complex in passive flexion; a single degree of freedom system. Journal of biomechanics 1999; 32(2): 111-118 60. Levin D, Nazarian LN, Miller TT, et al. Lateral Epicondylitis of the Elbow: US Findings1. Radiology 2005; 237(1):230

61. Lieber RL, Fridén J. Musculoskeletal balance of the human wrist elucidated using intraoperative laser diffraction. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology 1998; 8(2): 93-100

62. Lieber RL, Ljung BO, Fridén J. Intraoperative sarcomere length measurements reveal differential design of human wrist extensor muscles. The Journal of experimental biology 1997a; 200(Pt 1): 19-25 63. Lieber RL, Ljung BO, Fridén J. Sarcomere length in wrist extensor muscles Changes may provide insights into the etiology of chronic lateral epicondylitis. Acta Orthopaedica 1997b; 68(3): 249-254 64. Li G, Pierce JE, Herndon JH. A global optimization method for prediction of muscle forces of human musculoskeletal system. Journal of biomechanics 2006; 39(3): 522-529

65. Li Z, Yao SJ, Alini M, Stoddart MJ. Chondrogenesis of human bone marrow mesenchymal stem cells in fibrin-polyurethane composites is modulated by frequency and amplitude of dynamic compression and shear stress. Tissue engineering. Part A 2010; 16(2): 575-584

66. London JT, Kinematics of the elbow. The Journal of Bone and Joint Surgery 1981; 63(4):529 67. Lui PP, Chan LS, Lee YW, Fu SC, Chan KM. Sustained expression of proteoglycans and collagen type III/ type I ratio in a calcified tendinopathy model. Rheumatology (Oxford, England) 2010; 49(2): 231-239 68. Maganaris CN, Tensile properties of in vivo human tendinous tissue. Journal of biomechanics 2002; 35(8): 1019-1027 69. Maganaris CN, Paul JP. In vivo human tendinous tissue stretch upon maximum muscle force generation. Journal of biomechanics 2000; 33(11): 1453-1459

70. Magnusson SP, Langberg H, Kjaer M. The pathogenesis of tendinopathy: balancing the response to loading. Nature reviews. Rheumatology 2010; 6(5): 262-268

71. Magnusson SP, Narici MV, Maganaris CN, Kjaer M. Human tendon behaviour and adaptation, in vivo. The Journal of physiology 2008; 586(1): 71-81

72. Manchanda G, Grover D. Effectiveness of movement with mobilization compared with manipulation of wrist in case of lateral epicondylitis. Indian Journal of Physiotherapy and Occupational Therapy 2007; 16

73. Martin JA, Mehr D, Pardubsky PD, Buckwalter JA. The role of tenascin-C in adaptation of tendons to compressive loading. Biorheology 2003; 40(1-3): 321-329

74. Michna H, Morphometric analysis of loading-induced changes in collagen-fibril populations in young tendons. Cell and tissue research 1984; 236(2): 465-470

75. Milz S, Aktas T, Putz R, Benjamin M. Expression of extracellular matrix molecules typical of articular cartilage in the human scapholunate interosseous ligament. Journal of anatomy 2006; 208(6):671 76. Milz S, Benjamin M, Putz R. Molecular parameters indicating adaptation to mechanical stress in fibrous connective tissue. Advances in anatomy, embryology, and cell biology 2005; 178 1-71

77. Milz S, Tischer T, Buettner A, et al. Molecular composition and pathology of entheses on the medial and lateral epicondyles of the humerus: a structural basis for epicondylitis. Annals of the rheumatic diseases 2004; 63(9): 1015-1021

78. Montgomery SC, Moorehead JD, Davidson JS, Lowe D, Dangerfield PH. A new technique for measuring the rotational axis pathway of a moving knee. The Knee 1998; 5(4): 289-295

79. Moojen TM, Carpal Kinematics. Academisch Proefschrift Universiteit van Amsterdam 2003;

80. Moojen TM, Snel JG, Ritt M, Kauer JMG, Venema HW, Bos KE. Three-dimensional carpal kinematics in vivo. Clinical Biomechanics 2002a; 17(7): 506-514

81. Moojen TM, Snel JG, Ritt MJ, Venema HW, Kauer JM, Bos KE. Scaphoid kinematics in vivo. The Journal of hand surgery 2002b; 27(6): 1003-1010

82. Morree de JJ. Dynamiek van menselijk bindweefsel. Functie, beschadiging en herstel. Houten/Diegem: Bohn Stafleu Van Loghum; 2001.

83. Narici MV, Maffulli N, Maganaris CN. Ageing of human muscles and tendons. Disability and rehabilitation 2008; 30(20-22): 1548-1554

84. Narici MV, Maganaris CN. Plasticity of the muscle-tendon complex with disuse and aging. Exercise and sport sciences reviews 2007; 35(3): 126-134

85. Neu CP, Crisco JJ, Wolfe SW. In vivo kinematic behavior of the radio-capitate joint during wrist flexionextension and radio-ulnar deviation. Journal of biomechanics 2001; 34(11): 1429-1438

86. Newsham-West R, Nicholson H, Walton M, Milburn P. Long-term morphology of a healing bonetendon interface: a histological observation in the sheep model. Journal of anatomy 2007; 210(3): 318-327

87. Oonk H. Osteo- en Arthrokinematica. Weert: Henric Graaff van IJssel; 1988.

88. Ottani V, Martini D, Franchi M, Ruggeri A, Raspanti M. Hierarchical structures in fibrillar collagens. Micron 2002; 33(7-8): 587-596

89. Ottani V, Raspanti M, Ruggeri A. Collagen structure and functional implications. Micron 2001; 32(3): 251-260

90. Pierce JE, Li G. Muscle forces predicted using optimization methods are coordinate system dependent. Journal of biomechanics 2005; 38(4): 695-702

91. Puchner N, The effectiveness of manipulation of the wrist in the management of lateral epicondylitis. A dissertation. http://ujdigispace.uj.ac.za:8080/dspace/handle/10210/827 2008;

92. Radpasand M, Combination of manipulation, exercise, and physical therapy for the treatment of a 57year-old woman with lateral epicondylitis. Journal of manipulative and physiological therapeutics 2009; 32(2): 166-172

93. Reeves ND, Maganaris CN, Ferretti G, Narici MV. Influence of 90-day simulated microgravity on human tendon mechanical properties and the effect of resistive countermeasures. Journal of applied physiology (Bethesda, Md. : 1985) 2005; 98(6): 2278-2286

94. Riek S, Carson RG, Wright A. A new technique for the selective recording of extensor carpi radialis longus and brevis EMG. Journal of electromyography and kinesiology : official journal of the International Society of Electrophysiological Kinesiology 2000; 10(4): 249-253

95. Riezebos C, Lagerberg A. De tennispols. Versus, Tijdschrift voor Fysiotherapie 1988; 6(5): 228-243 96. Riley G, Tendinopathy--from basic science to treatment. Nature clinical practice. Rheumatology 2008; 4(2): 82-89

97. Scarvell JM, Smith PN, Refshauge KM, Galloway H, Woods K. Comparison of kinematics in the healthy and ACL injured knee using MRI. Journal of biomechanics 2005; 38(2): 255-262

98. Schomacher J, The convex-concave rule and the lever law. Manual therapy 2009; 14(5): 579-582 99. Screen HR, Chhaya VH, Greenwald SE, Bader DL, Lee DA, Shelton JC. The influence of swelling and matrix degradation on the microstructural integrity of tendon. Acta biomaterialia 2006; 2(5): 505-513 100. Screen HR, Shelton JC, Bader DL, Lee DA. Cyclic tensile strain upregulates collagen synthesis in isolated tendon fascicles. Biochemical and biophysical research communications 2005; 336(2): 424-429 101. Sears WR, McCombe PF, Sasso RC. Kinematics of cervical and lumbar total disc replacement. Semin Spine Surg 2006; 18 117-129

102. Seynnes OR, Erskine RM, Maganaris CN, et al. Training-induced changes in structural and mechanical properties of the patellar tendon are related to muscle hypertrophy but not to strength gains. Journal of applied physiology (Bethesda, Md. : 1985) 2009; 107(2): 523-530

103. Shetty A, Randolph M. A biomechanical analysis of injury, prevention, and rehabilitation exercises for lateral epicondylitis: a review. 18 International Symposium on Biomechanics in Sports, Hong Kong, China 2000;

104. Short WH, Werner FW, Green JK, Sutton LG, Brutus JP. Biomechanical evaluation of the ligamentous stabilizers of the scaphoid and lunate: part III. The Journal of hand surgery 2007; 32(3): 297-309 105. Slater H, Arendt-Nielsen L, Wright A, Graven-Nielsen T. Experimental deep tissue pain in wrist extensors--a model of lateral epicondylalgia. Eur J Pain 2003; 7(3): 277-288

106. Smedt de T, de Jong A, Van Leemput W, Lieven D, Van Glabbeek F. Lateral epicondylitis in tennis: update on aetiology, biomechanics and treatment. British journal of sports medicine 2007; 41(11): 816-819

107. Smidt N, van der Windt DAWM, Assendelft WJJ, Devillé WLJM, Korthals-de Bos IBC, Bouter LM. Corticosteroid injections, physiotherapy, or a wait-and-see policy for lateral epicondylitis: a randomised controlled trial. The lancet 2002; 359(9307): 657-662

108. Snijders CJ, Volkers ACW, Mechels K, Vleeming A. Provocation of epicondylagia lateralis (tennis elbow) by power grip pinching. Medicine and Science in Sports and Exercise 1987; 19(5): 518-523 109. Stasinopoulos D, Johnson MI. It may be time to modify the Cyriax treatment of lateral epicondylitis. Journal of Bodywork & Movement Therapies 2007; 11(1): 64-67

110. Stoeckart R, Vleeming A, Simons JL, van Helvoirt RP, Snijders CJ. Fascial deformation in the lateral elbow region: a conceptual approach. Clinical Biomechanics 1991; 6(1): 60-62

111. Stoeckart R, Vleeming A, Snijders CJ. Anatomy of the extensor carpi radialis brevis muscle related to tennis elbow. Clinical Biomechanics 1989; 4(4): 210-212

112. Struijs PA, Damen PJ, Bakker EW, Blankevoort L, Assendelft WJ, van Dijk CN. Manipulation of the wrist for management of lateral epicondylitis: a randomized pilot study. Physical therapy 2003; 83(7): 608-616 113. Sun JS, Shih TT, Ko CM, Chang CH, Hang YS, Hou SM. In vivo kinematic study of normal wrist motion: an ultrafast computed tomographic study. Clinical biomechanics (Bristol, Avon) 2000; 15(3): 212-216 114. Tang JB, Ryu J, Kish V. Scapholunate interosseous ligament sectioning adversely affects excursions of radial wrist extensor and flexor tendons. The Journal of hand surgery 1997; 22(4): 720-725

115. Tang JB, Ryu J, Omokawa S, Wearden S. Wrist kinetics after scapholunate dissociation: the effect of scapholunate interosseous ligament injury and persistent scapholunate gaps. Journal of orthopaedic research : official publication of the Orthopaedic Research Society 2002; 20(2): 215-221

116. Tasker T, Waugh W. Articular changes associated with internal derangement of the knee. The Journal of bone and joint surgery. British volume 1982; 64(4): 486-488

117. Thomopoulos S, Genin GM, Galatz LM. The development and morphogenesis of the tendon-to-bone insertion - what development can teach us about healing -. Journal of musculoskeletal & neuronal interactions 2010; 10(1): 35-45

118. Thomopoulos S, Marquez JP, Weinberger B, Birman V, Genin GM. Collagen fiber orientation at the tendon to bone insertion and its influence on stress concentrations. Journal of biomechanics 2006; 39(10): 1842-1851

119. Tournay A, Paillard J. Électromyographie des muscles radiaux à l'état normal. Révue Neurologie 1953; 89 277-279

120. Trudel D, Duley J, Zastrow I, Kerr EW, Davidson R, MacDermid JC. Rehabilitation for patients with lateral epicondylitis: a systematic review. Journal of hand therapy : official journal of the American Society of Hand Therapists 2004; 17(2): 243-266

121. Trudel G, Uhthoff HK. Contractures secondary to immobility: is the restriction articular or muscular? An experimental longitudinal study in the rat knee. Archives of physical medicine and rehabilitation 2000; 81(1): 6-13

122. Tsuzaki M, Yamauchi M, Banes AJ. Tendon collagens: extracellular matrix composition in shear stress and tensile components of flexor tendons. Connective tissue research 1993; 29(2): 141-152

123. Vicenzino B, Cleland JA, Bisset L. Joint manipulation in the management of lateral epicondylalgia: a clinical commentary. The Journal of manual & manipulative therapy 2007; 15(1): 50-56

124. Vicenzino B, Paungmali A, Buratowski S, Wright A. Specific manipulative therapy treatment for chronic lateral epicondylalgia produces uniquely characteristic hypoalgesia. Manual therapy 2001; 6(4): 205-212

125. Wang JH, Mechanobiology of tendon. Journal of biomechanics 2006; 39(9): 1563-1582 126. Weiler PJ, Bogoch ER. Kinematics of the distal radioulnar joint in rheumatoid arthritis: an in vivo study using centrode analysis. The Journal of hand surgery 1995; 20(6): 937-943

127. Werner FW, Short WH, Green JK. Changes in patterns of scaphoid and lunate motion during functional arcs of wrist motion induced by ligament division. The Journal of hand surgery 2005; 30(6): 1156-1160

128. Wolfe SW, Neu C, Crisco JJ. In vivo scaphoid, lunate, and capitate kinematics in flexion and in extension. The Journal of hand surgery 2000; 25(5): 860-869

129. Wong M, Carter DR. Articular cartilage functional histomorphology and mechanobiology: a research perspective. Bone 2003; 33(1): 1-13

130. Woo SL, Abramowitch SD, Kilger R, Liang R. Biomechanics of knee ligaments: injury, healing, and repair. Journal of biomechanics 2006; 39(1): 1-20

131. Woo SL, Gomez MA, Sites TJ, Newton PO, Orlando CA, Akeson WH. The biomechanical and morphological changes in the medial collateral ligament of the rabbit after immobilization and remobilization. The Journal of bone and joint surgery. American volume 1987; 69(8): 1200-1211 132. Wren TA, Beaupré GS, Carter DR. Tendon and ligament adaptation to exercise, immobilization, and

remobilization. Journal of rehabilitation research and development 2000; 37(2): 217-224

133. Yasuda K, Hayashi K. Changes in biomechanical properties of tendons and ligaments from joint disuse. Osteoarthritis and cartilage / OARS, Osteoarthritis Research Society 1999; 7(1): 122-129 134. Yoon JH, Halper J. Tendon proteoglycans: biochemistry and function. Journal of musculoskeletal & neuronal interactions 2005; 5(1): 22-34